Quality Notification - IMM 2017-06

Actie - 14 maart 2017

Elecsys AMH plus

Lancering van de Elecsys AMH plus assay ter vervanging van de Elecsys AMH assay.

Overzicht		
Systemen	cobas e 411 cobas e 601 cobas e 602 cobas e 801 MODULAR <i>ANALYTICS</i> <e></e>	
Reagentia Calibratoren Controles	Elecsys AMH – e411/e601/e602 Elecsys AMH – e801 Elecsys AMH CalSet PreciControl AMH	ref. 06 331 076 190 ref. 07 026 714 190 ref. 06 331 084 190 ref. 06 709 966 190
	Elecsys AMH Plus – e411/e601/e602 Elecsys AMH Plus – e801 Elecsys AMH Plus CalSet PreciControl AMH Plus	ref. 07 957 190 190 ref. 07 957 246 190 ref. 07 957 203 190 ref. 07 957 211 190





Roche introduceert een belangrijke uitbreiding van de intended use van haar Elecsys AMH assay: de Companion Diagnostics (CDx) claim in het kader van IVF-behandeling voor de 'bepaling van de individuele dagelijkse dosis follitropin delta van Ferring' (Andersen et al., Fertil Steril. 2017 Feb;107(2):387-396.e4).

Deze CDx claim en samenwerking met Ferring vereiste een bijkomende registratie door de autoriteiten zodat de Elecsys AMH plus als nieuwe assay geregistreerd en gelanceerd werd.

De huidige AMH assay zal uiterlijk juni 2017 volledig vervangen worden door de Elecsys AMH plus assay.

1) Overzicht van de wijzigingen

Inzake formulering, productie en standaardisatie, referentiewaarden, alsook prijs zijn de Elecsys AMH en Elecsys AMH plus assays identieke produkten.

Het verschil tussen beide assays omvat:

- 1) de bijkomende indicatie zoals vermeld in de bijsluiter van de Elecsys AMH plus assay: 'bepaling van de individuele dagelijkse dosis follitropin delta van Ferring'.
- 2) De Elecsys AMH plus bijsluiter vermeldt dat het AMH-resultaat bij gebruik voor de dosering van follitropin delta van Ferring, vergezeld moet worden van volgend statement: 'Deze AMH waarde in pmol/L werd bekomen met de Elecsys AMH plus assay en is geschikt voor de geïndividualiseerde dosering van follitropin delta van Ferring'.
- 3) Bijkomende aandacht is vereist om ervoor te zorgen dat de accuraatheid en precisie van de AMH plus test binnen de QC-limieten blijft
 - Target ranges van de AMH Plus controles
 - Interrun variatie =< 8% CV
 - Bias < +/-12%
 - Total errror < +/- 25%

2) Praktische informatie

Onderstaand vindt u een vergelijking van de Elecsys AMH en AMH plus assays:

EI	ecsys AMH
	Elecsys AMH
06 331 076 190	e411/e601/e602
07 026 714 490	e801
06 331 084 190	Elecsys AMH CalSet
06 709 966 190	PreciControl AMH

Elecsys AMH plus				
	Elecsys AMH Plus			
07 957 190 190	e411/e601/e602			
07 957 246 190	e801			
07 957 203 190	Elecsys AMH Plus CalSet			
07 957 211 190	PreciControl AMH Plus			

Please note

Results that are intended to be used for the dosing of the follitropin delta of Ferring must be accompanied by the following statement:

'This AMH value in pmol/L was generated with the Elecsys AMH Plus assay and is suitable for the individualized dosing of follitropin delta of Ferring.'

Intended Use

The determination of AMH is used for the assessment of ovarian reserve and the prediction of response to controlled ovarian stimulation (COS) in conjunction with other clinical and laboratory findings.

Test principle

Identical

Reagents - working solutions

Identical

The reagent RackPack is labelled AMH

Calibration

Identical

Quality Control

For quality control, use PreciControl AMH.

In addition, other suitable quality control material may be used.

Controls for the various concentration ranges should be run individually at least once every 24 hours when the test is in use, once per reagent kit, and following each calibration.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Follow the applicable government regulations and local guidelines for quality control.

Intended Use

The determination of AMH is used for the assessment of ovarian reserve and the prediction of response to controlled ovarian stimulation (COS) in conjunction with other clinical and laboratory findings.

In addition, the determination of AMH (in pmol/L) in combination with body weight is used for the establishment of the individual daily dose of the human recombinant follicle-stimulating hormone (rFSH) follitropin delta of Ferring in COS for the development of multiple follicles in women undergoing an assisted reproductive technology program.

Test principle

Identical

Reagents - working solutions

Identica

The reagent RackPack is labelled AMHP

Calibration

Identical

Quality Control

For quality control, use PreciControl AMH Plus.

Additional suitable quality control material may also be used in combination with PreciControl AMH Plus.

Special care needs to be taken to ensure that the accuracy and precision of the testing stays within acceptable limits. Besides meeting the PreciControl AMH Plus target ranges provided, the user needs to ensure that the systematic bias with respect to the assigned target values is within +/- 12%, the intermediate precision CV is <8% and the maximal total error is within +/- 25% (TE = bias + [1.65 x CV]). It is recommended to use quality control rule software.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Follow the applicable government regulations and local guidelines for quality control.

Limitations - interference Limitations - interference Identical Identical **Limits and ranges** Limits and ranges Identical Identical Expected values Expected values Identical Identical Use of AMH for the assessment of ovarian reserve Use of AMH for the assessment of ovarian reserve Identical Use of AMH for the prediction of hyper-response to Use of AMH for the prediction of hyper-response to controlled ovarian stimulation controlled ovarian stimulation Identical Identical Use of AMH for individual daily dose determination of follitropin delta of Ferring Follitropin delta is produced in a human cell line (PER.C6®) by recombinant DNA technology. Ferring is market authorisation holder of follitropin delta. The AMH concentration (in pmol/L) determined by the Elecsys AMH Plus assay in combination with body weight was validated for the individual daily dose determination of follitropin delta in controlled ovarian stimulation for the development of multiple follicles in women undergoing an assisted reproductive technology program such as in IVF or intracytoplpasmic sperm injection (ICS) cycle. The Elecsys AMH Plus assay was assessed exclusively for the individual dose determination of follitropin delta (Ferring). The AMH-based individualised dosing regimen of follitropin delta was validated in the prospective phase III clinical study ESTHER-1, a randomised, controlled, assessor-blind, parallel groups, multicenter, multinational trial comparing the efficacy and safety of follitropin delta with follitropin alfa (randomised 1:1 ratio). Women aged 18-40 years undergoing controlled ovarian stimulation for IVF or ICSI were enrolled and followed a GnRH agonist protocol. In this trial, 665 IVF/ICSI patients randomised to the follitropin delta group were treated with an individual daily dose of follitropin delta determined on the basis of their body weight and their AMH (in pmol/L) concentration measured with the Elecsys AMH Plus assay. The individual daily dose of follitropin delta was maintained throughout stimulation with no dose adjustments. Important information: Clinicians who want to administer follitropin delta must read and understand the current prescribing information of follitropin delta (Ferring) applicable in their country prior to administration of the drug; this document provides the details on the actual dosing regimen, the clinical efficacy and safety of the drug.

Specific performance data

Identical

Identical

Analytical specificity

Specific performance data

Identical

Identical

Analytical specificity

3) Bijlagen:

- Bijsluiter
- Fact Sheet

Wij danken u voor het vertrouwen dat u in ons stelt en blijven te uwer beschikking

Mocht u nog vragen hebben kunt u steeds terecht bij onze Regional Support

- op het nummer 02 247 45 16;
- via het e-mailadres: vilvoorde.support_nl@roche.com.

Alle informatie vindt u eveneens op onze website www.e-labdoc.be.

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Women's Health

Sabrina Schilling

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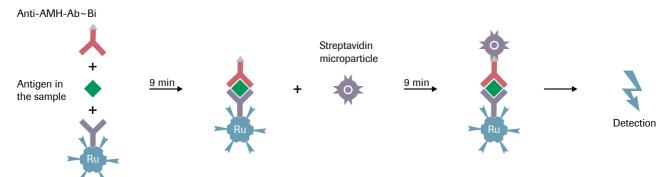
Electrochemiluminescence immunoassay (ECLIA) for the in vitro quantitative determination of anti-Müllerian Hormone in human serum and plasma

Indication

Immunoassay for the in vitro quantitative determination of anti-Müllerian hormone (AMH) in human serum and plasma. The determination of AMH is used for the assessment of the ovarian reserve and the prediction of response to controlled ovarian stimulation (COS) in conjunction with other clinical and laboratory findings. The Elecsys AMH Plus assay may also be used, in combination with body weight, to establish the individualised daily dose of the human recombinant follicle stimulating hormone (rFSH) follitropin delta of Ferring in women undergoing an assisted reproductive technology program.¹

AMH plays a fundamental role in the regression of Müllerian ducts in male embryo and in its absence, Müllerian ducts develop into female inner reproductive organs.² In females, it is secreted by the granulosa cells of pre-antral and small antral ovarian follicles. AMH regulates follicle recruitment and growth of small ovarian follicles while preventing exhaustion of follicular pool.^{3,4}

Test principle: Sandwich principle



Anti-AMH-Ab~Ru

1st Incubation (9 minutes):

50 μL of sample, a biotinylated monoclonal mammalian AMH-specific antibody and a monoclonal mammalian AMH-specific antibody labeled with a ruthenium complex react to form a sandwich complex.

2nd Incubation (9 minutes):

After addition of streptavidincoated microparticles, the complex becomes bound to the solid phase via interaction of biotin and streptavidin.

Measurement:

The reaction mixture is aspirated into the measuring cell where the microparticles are magnetically captured onto the surface of the electrode. Unbound substances are then removed. Application of a voltage to the electrode then induces chemiluminescent emission which is measured by a photomultiplier.

Results are determined via a calibration curve which is instrument-specifically generated by 2-point calibration and a master curve provided via the reagent barcode.





Elecsys® technology

ECL (ElectroChemiLuminescence) is a highly sensitive detection technology used with Roche immunoassays. Based on this technology and combined with well-designed, specific and sensitive immunoassays, Elecsys delivers reliable results. The development of ECL immunoassays is based on the use of a ruthenium-complex and tripropylamine (TPA). The chemiluminescence reaction for the detection of the reaction complex is initiated by applying a voltage to the sample solution resulting in a precisely controlled reaction. ECL technology can accommodate many immunoassay principles while providing superior performance.

Elecsys Anti-Müllerian Hormone (AMH Plus) test characteristics

Testing time	18 min
Test principle	Sandwich assay
Calibration	2 point
Traceability	Standardized against BCI AMH Gen II ELISA (unmodified)
Sample material	Serum and Li-heparin plasma
Sample volume	
LoB, LoD, LoQ*	0.049 pmol/L (0.007 ng/mL), 0.07 pmol/L (0.010 ng/mL), 0.21 pmol/L (0.030 ng/mL)
Measuring range	0.07 – 164 pmol/L (0.01 – 23 ng/mL)
Intermediate imprecision	cobas e 411 analyzer: 2.9 - 4.4 % cobas e 601/e 602 modules: 2.7 - 3.5 % Lowest conc. measured: 1.656 pmol/L (0.232 ng/mL)

Expected values

	N	2.5	th perc.	5 th	perc.	Me	dian	95 th	perc.	97.5 ^t	perc.
		pmol/L	ng/mL	pmol/L	ng/mL	pmol/L	ng/mL	pmol/L	ng/mL	pmol/L	ng/mL
Healthy r	men										
	148	5.50	0.77	10.2	1.43	34.2	4.79	82.8	11.6	103	14.5
Healthy v	women (y	ears)									
20 – 24	150	8.71	1.22	10.9	1.52	28.6	4.00	71.0	9.95	83.6	11.7
25 – 29	150	6.35	0.890	8.57	1.20	23.6	3.31	64.6	9.05	70.3	9.85
30 – 34	138	4.11	0.576	5.08	0.711	20.0	2.81	54.2	7.59	58.0	8.13
35 – 39	138	1.05	0.147	2.89	0.405	14.2	2.00	49.7	6.96	53.5	7.49
40 – 44	142	0.193	0.027	0.421	0.059	6.29	0.882	31.7	4.44	39.1	5.47
45 – 50	169	0.071	0.010	0.071	0.010	1.39	0.194	12.8	1.79	19.3	2.71
PCOS wo	omen**										
	149	13.3	1.86	17.2	2.41	48.6	6.81	122	17.1	135	18.9

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

- * LoB = Limit of Blank; LoD = Limit of Detection; LoQ = Limit of Quantitation (20 % total error)
- ** According to the revised diagnostic criteria of PCOS defined by the Rotterdam ESHRE/ASRM-sponsored (ESHRE = European Society of Human Reproduction and Embryology; ASRM = American Society of Reproductive Medicine) PCOS consensus workshop group⁵

Order information		
Elecsys AMH Plus	100 tests	07957190 190
Elecsys AMH Plus CalSet	4 x 1.0 mL	07957203 190
PreciControl AMH Plus	4 x 2.0 mL	07957211 190
Universal Diluent 2	2 x 36 ml	05192943 190

References

- 1 Anti-Müllerian Hormone. Elecsys and cobas e analyzers package insert 2016.
- 2 La Marca, A., Broekmans, F.J. (2009). *Hum Reprod*; 24(9):2264-75.
- 3 Visser, J.A., Themmen, A.P. (2005). Mol Cell Endocrinol; 234(1-2):81-6.
- 4 La Marca, A., Volpe, A. (2007). Hum Reprod Update; 13(3):265-73.
- 5 Rotterdam ESHRE/ASRM-Sponsored PCOS consensus workshop group (2004). Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome (PCOS). *Hum Reprod*; 19(1): 41-47.

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Elecsys AMH Plus



REF	\sum	SYSTEM
		MODULAR ANALYTICS E170
07957190 190	100	cobas e 411
		cobas e 601
		cobas e 602

English

System information

For **cobas e** 411 analyzer: test number 1590 For MODULAR ANALYTICS E170, **cobas e** 601 and **cobas e** 602 analyzers: Application Code Number 472

Please note

Results that are intended to be used for the dosing of the follitropin delta of Ferring must be accompanied by the following statement: This AMH value in pmol/L was generated with the Elecsys AMH Plus assay and is suitable for the individualized dosing of follitropin delta of Ferring.

Intended use

Immunoassay for the in vitro quantitative determination of anti-Müllerian hormone (AMH) in human serum and plasma. The determination of AMH is used for the assessment of the ovarian reserve and the prediction of response to controlled ovarian stimulation (COS) in conjunction with other clinical and laboratory findings.

In addition, the determination of AMH (in pmol/L) in combination with body weight is used for the establishment of the individual daily dose of the human recombinant follicle-stimulating hormone (rFSH) follitropin delta of Ferring (in accordance with the current prescribing information of the Ferring follitropin delta) in controlled ovarian stimulation for the development of multiple follicles in women undergoing an assisted reproductive technology program.

The **e**lectro**c**hemiluminescence **i**mmuno**a**ssay "ECLIA" is intended for use on Elecsys and **cobas e** immunoassay analyzers.

Summary

The anti-Müllerian hormone is a homodimeric glycoprotein belonging to the transforming growth factor β (TGF β) family. All members of this superfamily are involved in the regulation of tissue growth and differentiation. Prior to secretion, the hormone undergoes glycosylation and dimerization to produce an approximately 140 kDa precursor of two identical disulfide-linked 70 kDa subunits. Each monomer contains a large N-terminal proregion and a much smaller C-terminal mature domain. In contrast to other TGF β family members, AMH is thought to require the N-terminal domain to potentiate activity of the C-terminal domain to attain full bioactivity. $^{1.2}$

A part of AMH is then cleaved at a specific site between the pro-region and the mature region during cytoplasmic transit to generate biologically active 110 kDa N-terminal and 25 kDa C-terminal homodimers which remain associated in a non-covalent complex. The AMH type II receptor (AMH RII) has the capacity of binding only the biologically active form of AMH.²

In males, AMH is secreted by the Sertoli cells of the testes. During embryonic development in males, secretion of AMH from testicular Sertoli cells is responsible for the regression of the Müllerian duct and the normal development of the male reproductive tract. The secretion of AMH by the Sertoli cells starts during the embryogenesis and continues throughout life. AMH is continuously produced by the testicles until puberty and then decreases slowly to post-puberty values.³

In females AMH plays an important role in the ovarian folliculogenesis.⁴ Follicle development in the ovaries comprises two distinct stages: initial recruitment, by which primordial follicles start to mature, and cyclic recruitment, which leads to the growth of a cohort of small antral follicles, among which the dominant follicle (destined to ovulate) is subsequently selected. FSH directs the cyclic recruitment. AMH expression in granulosa cells starts in primary follicles and is maximal in granulosa cells of preantral and small antral follicles up to approximately 6 mm in diameter. When follicle growth becomes FSH-dependent, AMH expression diminishes and becomes undetectable. This pattern of AMH expression supports the inhibitory role of AMH at two distinct stages of folliculogenesis. First, AMH inhibits the transition of follicles from primordial into maturation stages and thereby has an important role in regulating the number of follicles remaining

in the primordial pool. Second, AMH has inhibitory effects on follicular sensitivity to FSH and therefore has a role in the process of follicular selection $^{5.6}$

Serum levels of AMH are barely detectable at birth in females, reach their highest levels after puberty, decrease progressively thereafter with age, and become undetectable at menopause. ^{7,8} Serum AMH levels have been shown to be relatively stable during the menstrual cycle with substantial fluctuations being observed in younger women. ^{9,10,11} AMH levels further demonstrate lower intra- and inter-cyclic variation than baseline FSH. ¹⁰ Serum AMH levels decrease significantly during the use of combined contraceptives. ¹² Clinical applications of AMH measurements have been proposed for a variety of indications. ^{13,14,15} Measurement of serum AMH is clinically mainly used for assessment of ovarian reserve reflecting the number of antral and pre-antral follicles, the so-called antral follicle count (AFC), and for the prediction of response to controlled ovarian stimulation. ^{13,15,16} Further clinical applications of AMH are diagnosis of disorders of sex development in children ^{17,18} and monitoring of granulosa cell tumors to detect residual or recurrent disease. ^{19,20} AMH has been suggested as a surrogate biomarker for AFC in the diagnosis of polycystic ovary syndrome (PCOS)^{21,22} and for the prediction of time to menopause. ^{23,24}

Test principle

Sandwich principle. Total duration of assay: 18 minutes.

- 1st incubation: 50 µL of sample, a biotinylated monoclonal AMH-specific antibody, and a monoclonal AMH-specific antibody labeled with a ruthenium complex^a) form a sandwich complex.
- 2nd incubation: After addition of streptavidin-coated microparticles, the complex becomes bound to the solid phase via interaction of biotin and streptavidin.
- The reaction mixture is aspirated into the measuring cell where the microparticles are magnetically captured onto the surface of the electrode. Unbound substances are then removed with ProCell/ProCell M. Application of a voltage to the electrode then induces chemiluminescent emission which is measured by a photomultiplier.
- Results are determined via a calibration curve which is instrumentspecifically generated by 2-point calibration and a master curve provided via the reagent barcode or e-barcode.
- a) Tris(2,2'-bipyridyl)ruthenium(II)-complex (Ru(bpy)₃²⁺)

Reagents - working solutions

The reagent rackpack is labeled as AMHP.

- M Streptavidin-coated microparticles (transparent cap), 1 bottle, 6.5 mL: Streptavidin-coated microparticles 0.72 mg/mL; preservative.
- R1 Anti-AMH-Ab~biotin (gray cap), 1 bottle, 8 mL:
 Biotinylated monoclonal anti-AMH antibody (mouse) 1.0 mg/L, phosphate buffer 50 mmol/L, pH 7.5; preservative.
- R2 Anti-AMH-Ab~Ru(bpy)₃²⁺ (black cap), 1 bottle, 8 mL:

 Monoclonal anti-AMH antibody (mouse) labeled with ruthenium complex 1.0 mg/L, phosphate buffer 50 mmol/L, pH 7.5; preservative.

Precautions and warnings

For in vitro diagnostic use.

Exercise the normal precautions required for handling all laboratory reacents

Disposal of all waste material should be in accordance with local guidelines. Safety data sheet available for professional user on request.

Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).



Reagent handling

The reagents in the kit have been assembled into a ready-for-use unit that cannot be separated.

All information required for correct operation is read in from the respective reagent barcodes.

Storage and stability

Store at 2-8 °C.

Do not freeze.

Store the Elecsys reagent kit **upright** in order to ensure complete availability of the microparticles during automatic mixing prior to use.

Stability:	
unopened at 2-8 °C	up to the stated expiration date
after opening at 2-8 °C	12 weeks
on the analyzers	8 weeks

Specimen collection and preparation

Only the specimens listed below were tested and found acceptable. Serum collected using standard sampling tubes or tubes containing separating gel.

Li-heparin plasma. Do not use EDTA plasma.

Criterion: Recovery within \pm 30 % of serum value \geq 3.57 pmol/L (\geq 0.5 ng/mL); recovery within \pm 1.43 pmol/L (\pm 0.2 ng/mL) for serum value < 3.57 pmol/L (< 0.5 ng/mL) and slope of 0.9-1.1 + intercept within \pm 0.7 pmol/L (\pm 0.1 ng/mL) + coefficient of correlation \geq 0.95.

Stable for 3 days at 20-25 °C, 5 days at 2-8 °C, 6 months at -20 °C (\pm 5 °C). Freeze only once.

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube

Centrifuge samples containing precipitates before performing the assay. Do not use heat-inactivated samples.

Do not use samples and controls stabilized with azide.

Ensure the samples, calibrators and controls are at 20-25 °C prior to measurement.

Due to possible evaporation effects, samples, calibrators and controls on the analyzers should be analyzed/measured within 2 hours.

Materials provided

See "Reagents – working solutions" section for reagents.

Materials required (but not provided)

- REF 07957203190, CalSet AMH Plus, for 4 x 1 mL
- REF 07957211190, PreciControl AMH Plus, for 2 x 2 mL each of PreciControl AMH Plus 1 and 2
- REF 05192943190, Diluent Universal 2, 2 x 36 mL sample diluent or
 REF 11732277122, Diluent Universal, 2 x 16 mL sample diluent or
 REF 03183971122, Diluent Universal, 2 x 36 mL sample diluent
- General laboratory equipment
- MODULAR ANALYTICS E170 or cobas e analyzer

Accessories for cobas e 411 analyzer:

- REF 11662988122, ProCell, 6 x 380 mL system buffer
- REF 11662970122, CleanCell, 6 x 380 mL measuring cell cleaning solution
- REF 11930346122, Elecsys SysWash, 1 x 500 mL washwater additive
- REF 11933159001, Adapter for SysClean
- REF 11706802001, AssayCup, 60 x 60 reaction cups
- REF 11706799001, AssayTip, 30 x 120 pipette tips
- REF 11800507001, Clean-Liner

Accessories for MODULAR ANALYTICS E170, cobas e 601 and cobas e 602 analyzers:

- REF 04880340190, ProCell M, 2 x 2 L system buffer
- REF 04880293190, CleanCell M, 2 x 2 L measuring cell cleaning solution
- REF 03023141001, PC/CC-Cups, 12 cups to prewarm ProCell M and CleanCell M before use
- REF 03005712190, ProbeWash M, 12 x 70 mL cleaning solution for run finalization and rinsing during reagent change
- REF 03004899190, PreClean M, 5 x 600 mL detection cleaning solution
- REF 12102137001, AssayTip/AssayCup, 48 magazines x 84 reaction cups or pipette tips, waste bags
- REF 03023150001, WasteLiner, waste bags
- REF 03027651001, SysClean Adapter M

Accessories for all analyzers:

 REF 11298500316, ISE Cleaning Solution/Elecsys SysClean, 5 x 100 mL system cleaning solution

Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

Resuspension of the microparticles takes place automatically prior to use. Read in the test-specific parameters via the reagent barcode. If in exceptional cases the barcode cannot be read, enter the 15-digit sequence of numbers (except for the **cobas e** 602 analyzer).

MODULAR ANALYTICS E170, cobas e 601 and cobas e 602 analyzers: PreClean M solution is necessary.

Bring the cooled reagents to approximately 20 °C and place on the reagent disk (20 °C) of the analyzer. Avoid foam formation. The system automatically regulates the temperature of the reagents and the opening/closing of the bottles.

Calibration

Traceability: This method has been standardized against the Beckman Coulter AMH Gen II ELISA (unmodified version without predilution) assay.

Every Elecsys reagent set has a barcoded label containing specific information for calibration of the particular reagent lot. The predefined master curve is adapted to the analyzer using the relevant CalSet.

Calibration frequency: Calibration must be performed once per reagent lot using fresh reagent (i.e. not more than 24 hours since the reagent kit was registered on the analyzer). Renewed calibration is recommended as follows:

- after 1 month (28 days) when using the same reagent lot
- after 7 days (when using the same reagent kit on the analyzer)
- as required: e.g. quality control findings outside the defined limits

Quality control

For quality control, use PreciControl AMH Plus.

Additional suitable quality control material may also be used in combination with PreciControl AMH Plus.

Controls for the various concentration ranges should be run individually at least once every 24 hours when the test is in use, once per reagent kit, and following each calibration.

Special care needs to be taken to ensure that the accuracy and precision of the testing stays within acceptable limits. Besides meeting the PreciControl AMH Plus target ranges provided, the user needs to ensure that the systematic bias with respect to the assigned target value is within \pm 12 %, the intermediate precision CV is \leq 8 % and the maximal total error is within \pm 25 % (TE = |biasl + 1.65*CV). It is recommended to use quality control rule software.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Follow the applicable government regulations and local guidelines for quality control.



Calculation

The analyzer automatically calculates the analyte concentration of each sample (either in pmol/L or in ng/mL).

Conversion factors: $pmol/L \times 0.14 = ng/mL$

 $ng/mL \times 7.14 = pmol/L$

Limitations - interference

The assay is unaffected by icterus (bilirubin \leq 1129 µmol/L or \leq 66 mg/dL), hemolysis (Hb \leq 0.621 mmol/L or \leq 1.0 g/dL), lipemia (Intralipid \leq 1000 mg/dL), biotin (\leq 143 nmol/L or \leq 30 ng/mL), lgG \leq 2.5 g/dL, lgA \leq 1.8 g/dL and lgM \leq 0.5 g/dL.

Criterion: Recovery within ± 10 % of initial value.

Samples should not be taken from patients receiving therapy with high biotin doses (i.e. > 5 mg/day) until at least 8 hours following the last biotin administration.

No interference was observed from rheumatoid factors up to a concentration of 1000 IU/mL.

There is no high-dose hook effect at AMH concentrations up to 9996 pmol/L (1400 ng/mL).

In vitro tests were performed on 20 commonly used pharmaceuticals. No interference with the assay was found up to the concentrations indicated within the below table.

Active Agent	Concentration tested mg/L
Acetylcysteine	150
Ampicillin-Na	1000
Ascorbic acid	300
Cyclosporine	5
Cefoxitin	2500
Heparin	5000 U
Levodopa	20
Methyldopa	20
Metronidazole	200
Phenylbutazone	400
Doxycycline	50
Acetylsalicylic acid	1000
Rifampicin	60
Acetaminophen	200
Ibuprofen	500
Theophylline	100
Triptorelin acetate	0.1
Metformin	2000
Folic Acid	0.4
Levothyroxine	0.2

In rare cases, interference due to extremely high titers of antibodies to analyte-specific antibodies, streptavidin or ruthenium can occur. These effects are minimized by suitable test design.

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

Limits and ranges

0.07-164 pmol/L (0.01-23 ng/mL) (defined by the Limit of Detection and the maximum of the master curve). Values below the Limit of Detection are reported as < 0.07 pmol/L (< 0.01 ng/mL). Values above the measuring range are reported as > 164 pmol/L (> 23 ng/mL) or up to 328 pmol/L (46 ng/mL) for 2-fold diluted samples.

Lower limits of measurement

Limit of Blank, Limit of Detection and Limit of Quantitation

Limit of Blank = 0.049 pmol/L (0.007 ng/mL)

Limit of Detection = 0.07 pmol/L (0.01 ng/mL)

Limit of Quantitation = 0.21 pmol/L (0.030 ng/mL)

The Limit of Blank, Limit of Detection and Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A2 requirements.

The Limit of Blank is the 95th percentile value from $n \ge 60$ measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95 %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples. The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

The Limit of Quantitation is the lowest analyte concentration that can be reproducibly measured with an intermediate precision CV of \leq 20 %.

Dilution

Samples with AMH concentrations above the measuring range can be diluted automatically with Diluent Universal 2. Manual dilution can be performed with Diluent Universal 2 or Diluent Universal. The recommended dilution is 1:2 (either automatically by the analyzers or manually). The concentration of the diluted sample must be $> 71.4 \, \text{pmol/L}$ ($> 10 \, \text{ng/mL}$).

After manual dilution, multiply the result by the dilution factor.

After dilution by the analyzers, the software automatically takes the dilution into account when calculating the sample concentration.

Expected values

A study in a Caucasian population with the Elecsys AMH Plus assay on samples from apparently healthy adults (148 males, 887 females not taking contraceptives) and 149 women with PCOS yielded the following results (Roche study No. RD001727):

	N	2.5th perc.	5 th perc.	Median	95th perc.	97.5 th
		ng/mL	ng/mL	ng/mL	ng/mL	perc.
		(95 % CI ^{b)})	(95 % CI)	(95 % CI)	(95 % CI)	ng/mL
						(95 % CI)
Healthy	men				•	1
	148	0.77	1.43	4.79	11.6	14.5
		(0.17-1.58)	(0.256-1.97)	(4.35-5.35)	(10.3-17.0)	(10.9-17.6)
Healthy	wome	en (years)				l
• 20-24	150	1.22	1.52	4.00	9.95	11.7
		(0.478-1.67)	(0.758-1.81)	(3.60-4.44)	(7.87-13.6)	(9.11-15.7)
• 25-29	150	0.890	1.20	3.31	9.05	9.85
		(0.493-1.21)	(0.797-1.75)	(3.00-3.89)	(7.59-10.3)	(8.91-11.3)
• 30-34	138	0.576	0.711	2.81	7.59	8.13
		(0.256-0.958)	(0.256-1.12)	(2.35-3.47)	(6.84-9.52)	(7.27-9.72)
• 35-39	138	0.147	0.405	2.00	6.96	7.49
		(0.053-0.474)	(0.053-0.496)	(1.73-2.36)	(5.31-9.37)	(6.49-10.9)
• 40-44	142	0.027	0.059	0.882	4.44	5.47
		(0.010-0.063)	(0.017-0.119)	(0.726-1.13)	(2.94-5.56)	(3.92-6.76)
• 45-50	169	0.010	0.010	0.194	1.79	2.71
		(0.010-0.010)	(0.010-0.010)	(0.144-0.269)	(1.43-2.99)	(1.79-4.16)
PCOS w	vomer)*			L	L
	149	1.86	2.41	6.81	17.1	18.9
		(1.54-2.50)	(1.67-3.01)	(6.30-7.42)	(13.3-20.3)	(16.0-21.1)

b) CI = confidence interval



	N	2.5th perc.	5 th perc.	Median	95 th perc.	97.5 th	
		pmol/L	pmol/L	pmol/L	pmol/L	perc.	
		(95 % CI)	(95 % CI)	(95 % CI)	(95 % CI)	pmol/L	
						(95 % CI)	
Healthy	Healthy men						
	148	5.5	10.2	34.2	82.8	103	
		(1.2-11.3)	(1.8-14.1)	(31.1-38.2)	(73.5-121)	(78.1-125)	
Healthy	wome	en (years)			•		
• 20-24	150	8.71	10.9	28.6	71.0	83.6	
		(3.41-11.9)	(5.41-12.9)	(25.7-31.7)	(56.2-97.1)	(65.0-112)	
• 25-29	150	6.35	8.57	23.6	64.6	70.3	
		(3.52-8.64)	(5.69-12.5)	(21.4-27.8)	(54.2-73.5)	(63.6-81.0)	
• 30-34	138	4.11	5.08	20.0	54.2	58.0	
		(1.83-6.84)	(1.83-8.00)	(16.8-24.8)	(48.8-68.0)	(51.9-69.4)	
• 35-39	138	1.05	2.89	14.2	49.7	53.5	
		(0.378-3.38)	(0.378-3.54)	(12.4-16.9)	(37.9-66.9)	(46.3-77.9)	
• 40-44	142	0.193	0.421	6.29	31.7	39.1	
		(0.071-0.450)	(0.121-0.850)	(5.18-8.07)	(21.0-39.7)	(28.0-48.3)	
• 45-50	169	0.071	0.071	1.39	12.8	19.3	
		(0.071-0.071)	(0.071-0.071)	(1.03-1.92)	(10.2-21.3)	(12.8-29.7)	
PCOS w	omer/)*		L	1	1	
	149	13.3	17.2	48.6	122	135	
		(11.0-17.8)	(11.9-21.5)	(45.0-53.0)	(95.0-145)	(114-151)	

^{*} According to the revised diagnostic criteria of PCOS defined by the Rotterdam ESHRE/ASRM-sponsored (ESHRE = European Society of Human Reproduction and Embryology; ASRM = American Society of Reproductive Medicine) PCOS consensus workshop group.²⁵

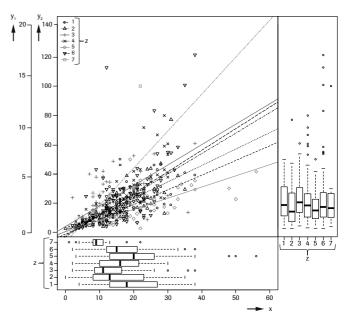
Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

Use of AMH for the assessment of ovarian reserve

The use of AMH for the assessment of ovarian reserve was investigated in a prospective study with n=451 women between 18-44 years old, where AMH values were correlated to the AFC of the women (Roche study No. RD001542). AFC was determined by transvaginal sonography measuring follicles of 2-10 mm diameter in size. Both AFC and AMH were determined on days 2-4 of the same menstrual cycle. Between 17 to 115 women were recruited per site at 6 different European sites and 1 Australian site.

No significant differences in mean AMH values were observed between the sites (pval = 0.301). The mean age values between the sites were significantly different, and also AMH and age showed a significant negative correlation (Spearman correlation coefficient -0.47). The age adjusted site effect of AMH showed no significance (pval = 0.193). The determined AFC values showed significant differences between the sites, with and without age adjustment. The overall correlation of AMH with AFC was 0.68 (Spearman's rank coefficient).

The figure below shows the scatterplot of AMH versus AFC, as well as the site-specific AMH and AFC distributions.



x: AFC (N)

y₁: AMH (ng/mL)

y₂: AMH (pmol/L)

z: Site

Agreement table on absolute AFC numbers of 7 and 15

Three AFC groups were defined 26,27 based on two cutoffs for AFC: 7 and 15 (0-7, 8-15, > 15). According to the prevalences within these groups (15 %, 37 %, 48 %), quantiles on AMH were computed (c1 = 4.86 pmol/L or 0.681 ng/mL, c2 = 16.2 pmol/L or 2.27 ng/mL) to define three groups. Agreement is presented in absolute numbers and percentages per AMH group.

Given the large variability of AFC results depending on site- and sonographer-specific variations, each site should review the agreement table for transferability to their own specific conditions.

	AFC 0-7	AFC 8-15	AFC > 15	N
AMH ≤ 4.86 pmol/L (0.681 ng/mL)	43 (63.2 %)	22 (32.4 %)	3 (4.4 %)	68
4.86 pmol/L (0.681 ng/mL) < AMH ≤ 16.2 pmol/L (2.27 ng/mL)	20 (12.0 %)	95 (56.9 %)	52 (31.1 %)	167
AMH > 16.2 pmol/L (2.27 ng/mL)	3 (1.4 %)	52 (24.1 %)	161 (74.5 %)	216
N	66	169	216	451

For a patient with AMH \leq 0.681 ng/mL the probability to have a low AFC (0-7) is 63 %, the probability to be in the middle AFC (8-15) group is about 32 % and only 4.4 % for having an AFC > 15.

The probability for patients with high AMH values (> 2.27 ng/mL) to have an AFC > 15 is 75 %, the probability for being in the middle AFC (8-15) group is 24 % and only 1.4 % to have an AFC < 8.

Use of AMH for the prediction of hyper-response to controlled ovarian stimulation

The following results were obtained in an external study "Clinical evaluation of the Elecsys AMH assay for the prediction of response to controlled ovarian stimulation" (Roche study No. CIM RD 001695).

AMH was determined in 149 women undergoing an antagonist treatment protocol in the course of their first cycle of controlled ovarian stimulation for in-vitro fertilization (IVF). Women included in the study were aged < 44 years, had a regular menstruation cycle and no major abnormalities at transvaginal sonography. Women with PCOS, endocrine or metabolic abnormalities and women undergoing IVF with oocyte donation were not



included. All women received a standard FSH stimulation dose of 150 IU/day. Blood was drawn before start of FSH stimulation for post hoc analysis of AMH after completion of the treatment cycle. Hyper-response was observed in 16 women. Hyper-response was defined as > 15 oocytes retrieved or cancellation of stimulation cycle where more than 20 follicles > 12 mm were observed and Estradiol values > 11700 pmol/L, or when more than 30 follicles > 12 mm were observed. The clinical performance of Elecsys AMH to predict hyper-response to controlled ovarian stimulation was evaluated by ROC (receiver operating characteristic) analysis and by applying a cutoff of 15 pmol/L (2.1 ng/mL) which has been previously published. ^{28,29} Prediction of hyper-response was significant with an AUC (area under the curve) of 82.1 % (Cl 72.5-91.7 %). Sensitivity, specificity, positive predictive value (PPV), and negative predictive value (NPV) for the AMH cutoff of 15.0 pmol/L (2.1 ng/mL) are shown in the table below.

Hyper-response				
AMH cutoff	15.0 pmol/L (2.10 ng/mL)			
	Estimate	95 % CI		
Sensitivity	81.3 %	54.4-96.0 %		
Specificity	64.7 %	55.9-72.8 %		
PPV	21.7 %	12.1-34.2 %		
NPV	96.6 %	90.5-99.3 %		

Use of AMH for individual daily dose determination of follitropin delta of Ferring

Follitropin delta is produced in a human cell line (PER.C6®) by recombinant DNA technology. ^{29,30,31} Ferring is marketing authorisation holder of follitropin delta.

The AMH concentration (in pmol/L) determined by the Elecsys AMH Plus assay in combination with body weight was validated for the individual daily dose determination of follitropin delta in controlled ovarian stimulation for the development of multiple follicles in women undergoing an assisted reproductive technology program such as an in IVF or intracytoplasmic sperm injection (ICSI) cycle. The Elecsys AMH Plus assay was assessed exclusively for the individual dose determination of follitropin delta (Ferring). The AMH-based individualised dosing regimen of follitropin delta was validated in the prospective phase III clinical study ESTHER-1, a randomised, controlled, assessor-blind, parallel groups, multicenter, multinational trial comparing the efficacy and safety of follitropin delta with follitropin alfa (randomized in 1:1 ratio). Women aged 18-40 years undergoing controlled ovarian stimulation for IVF or ICSI were enrolled and followed a GnRH antagonist protocol.³² In this trial, 665 IVF/ICSI patients randomised to the follitropin delta group were treated with an individual dose of follitropin delta determined on the basis of their body weight and their AMH (in pmol/L) concentration measured with the Elecsys AMH Plus assay. The individual daily dose of follitropin delta was maintained throughout stimulation with no dose adjustments.

Important information: Clinicians who want to administer follitropin delta must read and understand the current prescribing information of follitropin delta (Ferring) applicable in their country prior to administration of the drug; this document provides the details on the actual dosing regimen, the clinical efficacy and safety of the drug.³³

Specific performance data

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

Precision

Precision was determined using Elecsys reagents, samples and controls in a protocol (EP5-A2) of the CLSI (Clinical and Laboratory Standards Institute): 2 runs per day in duplicate each for 21 days (n = 84). The following results were obtained:

cobas e 411 analyzer					
		Repeatab	oility	Intermed precision	
Sample	Mean ng/mL	SD ng/mL	CV %	SD ng/mL	CV %
Human serum 1	0.232	0.004	1.8	0.010	4.4
Human serum 2	0.705	0.008	1.1	0.028	4.0

cobas e 411 analyzer					
		Repeatal	oility	Intermed precision	
Sample	Mean ng/mL	SD ng/mL	CV %	SD ng/mL	CV %
Human serum 3	2.44	0.030	1.2	0.082	3.3
Human serum 4	12.3	0.132	1.1	0.449	3.7
Human serum 5	18.8	0.287	1.5	0.711	3.8
PreciControl AMH 1	1.15	0.011	1.0	0.033	2.9
PreciControl AMH 2	5.68	0.057	1.0	0.214	3.8

MODULAR ANALYTICS E170, cobas e 601 and cobas e 602 analyzers					
		Repeatab	oility	Intermed precision	
Sample	Mean ng/mL	SD ng/mL	CV %	SD ng/mL	CV %
Human serum 1	0.248	0.004	1.7	0.008	3.2
Human serum 2	0.745	0.009	1.2	0.020	2.7
Human serum 3	2.55	0.028	1.1	0.070	2.7
Human serum 4	13.0	0.165	1.3	0.397	3.0
Human serum 5	19.6	0.188	1.0	0.650	3.3
PreciControl AMH 1	1.19	0.014	1.2	0.042	3.5
PreciControl AMH 2	5.89	0.055	0.9	0.200	3.4

cobas e 411 analyzer					
		Repeatab	oility	Intermed precision	
Sample	Mean pmol/L	SD pmol/L	CV %	SD pmol/L	CV %
Human serum 1	1.66	0.030	1.8	0.072	4.4
Human serum 2	5.03	0.054	1.1	0.200	4.0
Human serum 3	17.4	0.213	1.2	0.584	3.3
Human serum 4	87.5	0.941	1.1	3.20	3.7
Human serum 5	134	2.05	1.5	5.08	3.8
PreciControl AMH 1	8.19	0.082	1.0	0.23	2.9
PreciControl AMH 2	40.5	0.405	1.0	1.53	3.8

MODULAR ANALYTICS E170, cobas e 601 and cobas e 602 analyzers					
		Repeatal	oility	Intermed precision	
Sample	Mean pmol/L	SD pmol/L	CV %	SD pmol/L	CV %
Human serum 1	1.77	0.030	1.7	0.057	3.2
Human serum 2	5.32	0.065	1.2	0.142	2.7
Human serum 3	18.2	0.197	1.1	0.497	2.7
Human serum 4	93.1	1.18	1.3	2.84	3.0
Human serum 5	140	1.34	1.0	4.64	3.3
PreciControl AMH 1	8.49	0.099	1.2	0.300	3.5
PreciControl AMH 2	42.0	0.392	0.9	1.43	3.4

Method comparison

a) A comparison of the Elecsys AMH Plus assay (y) with a commercially available AMH assay (x) using clinical samples gave the following correlations (ng/mL):

Number of samples measured: 548



Passing/Bablok³⁴ Linear regression y = 0.639x + 0.247 y = 0.592x + 0.541 y = 0.919 z = 0.976

The sample concentrations were between 0.046 and 16.5 ng/mL.

b) A comparison of the Elecsys AMH Plus assay (y) with a commercially available AMH assay (x) using clinical samples in the normal range up to 4 ng/mL gave the following correlations (ng/mL):

Number of samples measured: 340

Passing/Bablok³⁴ Linear regression y = 0.788x + 0.095 y = 0.720x + 0.206t = 0.884 r = 0.966

The sample concentrations were between 0.046 and 3.99 ng/mL.

Analytical specificity

The monoclonal antibodies used are highly specific to human AMH. The following cross-reactivities were found:

Cross-reactant	Concentration tested	Cross-reactivity %
Inhibin A	100 ng/mL	n. d. ^{c)}
Activin A	100 ng/mL	n. d.
LH	500 mIU/mL	n. d.
FSH	500 mIU/mL	n. d.

c) n. d. = not detectable

References

- Wilson CA, di Clemente N, Ehrenfels C, et al. Mullerian inhibiting substance requires its N-terminal domain for maintenance of biological activity, a novel finding within the transforming growth factor-beta superfamily. Mol Endocrinol 1993;7(2):247-257.
- 2 di Clemente N, Jamin SP, Lugovskoy A, et al. Processing of antimullerian hormone regulates receptor activation by a mechanism distinct from TGF-beta. Mol Endocrinol 2010;24(11):2193-206.
- Grinspon RP, Rey RA. Anti-müllerian hormone and sertoli cell function in paediatric male hypogonadism. Horm Res Paediatr 2010;73(2):81-92.
- 4 Broekmans FJ, Visser JA, Laven JS, et al. Anti-Müllerian hormone and ovarian dysfunction. Trends Endocrinol Metab 2008;19(9):340-347.
- Visser JA, de Jong FH, Laven JS, et al. Anti-Müllerian hormone: a new marker for ovarian function. Reproduction 2006;131(1):1-9.
- 6 Visser JA, Schipper I, Laven JS, et al. Anti-Müllerian hormone: an ovarian reserve marker in primary ovarian insufficiency. Nat Rev Endocrinol 2012;8(6):331-341.
- 7 Kelsey TW, Anderson RA, Wright P, et al. Data-driven assessment of the human ovarian reserve. Mol Hum Reprod 2012;18(2):79-87.
- 8 Nelson SM, Iliodromiti S, Fleming R, et al. Reference range for the antimüllerian hormone Generation II assay: a population study of 10,984 women, with comparison to the established Diagnostics Systems Laboratory nomogram. Fertil Steril 2014;101(2):523-529.
- 9 Tsepelidis S, Devreker F, Demeestere I, et al. Stable serum levels of anti-Mullerian hormone during the menstrual cycle: a prospective study in normo-ovulatory women. Hum Reprod 2007;22:1837–1840.
- 10 van Disseldorp J, Lambalk CB, Kwee J, et al. Comparison of inter- and intra-cycle variability of anti-Mullerian hormone and antral follicle counts. Hum Reprod 2010;25(1):221-227.
- 11 Overbeek A, Broekmans FJ, Hehenkamp WJ, et al. Intra-cycle fluctuations of anti-Müllerian hormone in normal women with a regular cycle: a re-analysis. Reprod Biomed Online 2012;24(6):664-669.
- 12 Kallio S, Puurunen J, Ruokonen A, et al. Antimüllerian hormone levels decrease in women using combined contraception independently of administration route. Fertil Steril 2013;99(5):1305-1310.
- 13 Nelson SM. Biomarkers of ovarian response: current and future applications. Fertil Steril 2013;99(4):963-969.

- 14 Anderson RA, Wallace WH. Antimüllerian hormone, the assessment of the ovarian reserve, and the reproductive outcome of the young patient with cancer. FertilSteril 2013;99(6):1469-1475.
- 15 Dewailly D, Andersen CY, Balen A, et al. The physiology and clinical utility of anti-Mullerian hormone in women. Hum Reprod Update 2014;20(3):370-385.
- 16 La Marca A, Sunkara SK. Individualization of controlled ovarian stimulation in IVF using ovarian reserve markers: from theory to practice. Hum Reprod Update 2014;20(1):124-140.
- 17 Josso N, Rey R, Picard JY. Testicular anti-Müllerian hormone: clinical applications in DSD. Semin Reprod Med 2012;30(5):364-373.
- 18 Rey RA, Grinspon RP, Gottlieb S, et al. Male hypogonadism: an extended classification based on a developmental, endocrine physiology-based approach. Andrology 2013;1(1):3-16.
- 19 La Marca A, Volpe A. The Anti-Mullerian hormone and ovarian cancer. Hum Reprod Update 2007;13(3):265-273.
- 20 Geerts I, Vergote I, Neven P, et al. The role of inhibins B and antimüllerian hormone for diagnosis and follow-up of granulosa cell tumors. Int J Gynecol Cancer 2009;19(5):847-855.
- 21 Dewailly D, Gronier H, Poncelet E, et al. Diagnosis of polycystic ovary syndrome (PCOS): revisiting the threshold values of follicle count on ultrasound and of the serum AMH level for the definition of polycystic ovaries. Hum Reprod 2011;26(11):3123-3129.
- 22 Iliodromiti S, Kelsey TW, Anderson RA, et al. Can anti-Mullerian hormone predict the diagnosis of polycystic ovary syndrome? A systematic review and meta-analysis of extracted data. J Clin Endocrinol Metab 2013;98(8):3332-3340.
- 23 Tehrani FR, Solaymani-Dodaran M, Tohidi M, et al. Modeling age at menopause using serum concentration of anti-mullerian hormone. J Clin Endocrinol Metab 2013;98(2):729-735.
- 24 Dólleman M, Depmann M, Eijkemans MJ, et al. Anti-Mullerian hormone is a more accurate predictor of individual time to menopause than mother's age at menopause. Hum Reprod 2014;29(3):584-591.
- 25 Rotterdam ESHRE/ASRM-Sponsored PCOS consensus workshop group. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome (PCOS). Hum Reprod 2004;19(1):41-47.
- 26 Ferraretti AP, La Marca A, Fauser BC, et al. ESHRE working group on Poor Ovarian Response Definition. ESHRE consensus on the definition of 'poor response' to ovarian stimulation for in vitro fertilization: the Bologna criteria. Hum Reprod 2011;26(7):1616-1624.
- 27 van Tilborg TC, Eijkemans MJ, Laven JS, et al. The OPTIMIST study: optimisation of cost effectiveness through individualised FSH stimulation dosages for IVF treatment. A randomised controlled trial. BMC Womens Health 2012;18;12:29.
- 28 Nelson SM, Yates RW, Fleming R. Serum anti-Müllerian hormone and FSH: prediction of live birth and extremes of response in stimulated cycles--implications for individualization of therapy. Hum Reprod 2007;22(9):2414-21.
- 29 Arce JC, Andersen AN, Fernández-Sánchez M, et al. Ovarian response to recombinant human follicle-stimulating hormone: a randomized, antimüllerian hormone-stratified, dose-response trial in women undergoing in vitro fertilization/intracytoplasmic sperm injection. Fertil Steril 2014;102(6):1633-40.e5.
- 30 Olsson H, Sandström R, Grundemar L. Different pharmacokinetic and pharmacodynamic properties of recombinant follicle-stimulating hormone (rFSH) derived from a human cell line compared with rFSH from a non-human cell line. J Clin Pharmacol. 2014;54(11):1299-307.
- 31 Bosch E, Nyboe Andersen A, Barri P, et al. Follicular and endocrine dose responses according to anti-Müllerian hormone levels in IVF patients treated with a novel human recombinant FSH (FE 999049). Clin Endocrinol (Oxf). 2015;83(6):902-12.
- 32 ClinicalTrials.gov Evidence-based Stimulation Trial with Human rFSH in Europe and Rest of World 1 (ESTHER-1). ClinicalTrials.gov Identifier: NCT01956110. Available at: http://clinicaltrials.gov/ct2/show/NCT01956110?term=NCT01956110.
- 33 Applicable prescribing information of follitropin delta; Ferring.



REF		\sum	SYSTEM
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English

System information

Short name	ACN (application code number)
AMHP	10158

Please note

Results that are intended to be used for the dosing of the follitropin delta of Ferring must be accompanied by the following statement: This AMH value in pmol/L was generated with the Elecsys AMH Plus assay and is suitable for the individualized dosing of follitropin delta of Ferring.

Intended use

Immunoassay for the in vitro quantitative determination of anti-Müllerian hormone (AMH) in human serum and plasma. The determination of AMH is used for the assessment of the ovarian reserve and the prediction of response to controlled ovarian stimulation (COS) in conjunction with other clinical and laboratory findings.

In addition, the determination of AMH (in pmol/L) in combination with body weight is used for the establishment of the individual daily dose of the human recombinant follicle-stimulating hormone (rFSH) follitropin delta of Ferring (in accordance with the current prescribing information of the Ferring follitropin delta) in controlled ovarian stimulation for the development of multiple follicles in women undergoing an assisted reproductive technology program.

The **e**lectro**c**hemiluminescence **i**mmuno**a**ssay "ECLIA" is intended for use on the **cobas e** 801 immunoassay analyzer.

Summary

The anti-Müllerian hormone is a homodimeric glycoprotein belonging to the transforming growth factor β (TGF β) family. All members of this superfamily are involved in the regulation of tissue growth and differentiation. Prior to secretion, the hormone undergoes glycosylation and dimerization to produce an approximately 140 kDa precursor of two identical disulfide-linked 70 kDa subunits. Each monomer contains a large N-terminal proregion and a much smaller C-terminal mature domain. In contrast to other TGF β family members, AMH is thought to require the N-terminal domain to potentiate activity of the C-terminal domain to attain full bioactivity. $^{1.2}$

A part of AMH is then cleaved at a specific site between the pro-region and the mature region during cytoplasmic transit to generate biologically active 110 kDa N-terminal and 25 kDa C-terminal homodimers which remain associated in a non-covalent complex. The AMH type II receptor (AMH RII) has the capacity of binding only the biologically active form of AMH.²

In males, AMH is secreted by the Sertoli cells of the testes. During embryonic development in males, secretion of AMH from testicular Sertoli cells is responsible for the regression of the Müllerian duct and the normal development of the male reproductive tract. The secretion of AMH by the Sertoli cells starts during the embryogenesis and continues throughout life. AMH is continuously produced by the testicles until puberty and then decreases slowly to post-puberty values.³

In females AMH plays an important role in the ovarian folliculogenesis. ⁴ Follicle development in the ovaries comprises two distinct stages: initial recruitment, by which primordial follicles start to mature, and cyclic recruitment, which leads to the growth of a cohort of small antral follicles, among which the dominant follicle (destined to ovulate) is subsequently selected. FSH directs the cyclic recruitment. AMH expression in granulosa cells starts in primary follicles and is maximal in granulosa cells of preantral and small antral follicles up to approximately 6 mm in diameter. When follicle growth becomes FSH-dependent, AMH expression diminishes and becomes undetectable. This pattern of AMH expression supports the inhibitory role of AMH at two distinct stages of folliculogenesis. First, AMH inhibits the transition of follicles from primordial into maturation stages and thereby has an important role in regulating the number of follicles remaining in the primordial pool. Second, AMH has inhibitory effects on follicular sensitivity to FSH and therefore has a role in the process of follicular selection. ^{5,6}

Serum levels of AMH are barely detectable at birth in females, reach their highest levels after puberty, decrease progressively thereafter with age, and become undetectable at menopause. ^{7,8} Serum AMH levels have been shown to be relatively stable during the menstrual cycle with substantial fluctuations being observed in younger women. ^{9,10,11} AMH levels further demonstrate lower intra- and inter-cyclic variation than baseline FSH. ¹⁰ Serum AMH levels decrease significantly during the use of combined contraceptives. ¹² Clinical applications of AMH measurements have been proposed for a variety of indications. ^{13,14,15} Measurement of serum AMH is clinically mainly used for assessment of ovarian reserve reflecting the number of antral and pre-antral follicles, the so-called antral follicle count (AFC), and for the prediction of response to controlled ovarian stimulation. ^{13,15,16} Further clinical applications of AMH are diagnosis of disorders of sex development in children ^{17,18} and monitoring of granulosa cell tumors to detect residual or recurrent disease. ^{19,20} AMH has been suggested as a surrogate biomarker for AFC in the diagnosis of polycystic ovary syndrome (PCOS)^{21,22} and for the prediction of time to menopause. ^{23,24}

Test principle

Sandwich principle. Total duration of assay: 18 minutes.

- 1st incubation: 30 µL of sample, a biotinylated monoclonal AMH-specific antibody, and a monoclonal AMH-specific antibody labeled with a ruthenium complex^a) form a sandwich complex.
- 2nd incubation: After addition of streptavidin-coated microparticles, the complex becomes bound to the solid phase via interaction of biotin and streptavidin.
- The reaction mixture is aspirated into the measuring cell where the microparticles are magnetically captured onto the surface of the electrode. Unbound substances are then removed with ProCell II M. Application of a voltage to the electrode then induces chemiluminescent emission which is measured by a photomultiplier.
- Results are determined via a calibration curve which is instrumentspecifically generated by 2-point calibration and a master curve provided via the cobas link.

a) Tris(2,2'-bipyridyl)ruthenium(II)-complex (Ru(bpy)3+3)

Reagents - working solutions

The **cobas e** pack is labeled as AMHP.

- M Streptavidin-coated microparticles, 1 bottle, 5.8 mL: Streptavidin-coated microparticles 0.72 mg/mL; preservative.
- R1 Anti-AMH-Ab~biotin, 1 bottle, 7.2 mL: Biotinylated monoclonal anti-AMH antibody (mouse) 1.0 mg/L; phosphate buffer 50 mmol/L, pH 7.5; preservative.
- R2 Anti-AMH-Ab~Ru(bpy)²⁺₃, 1 bottle, 7.2 mL: Monoclonal anti-AMH antibody (mouse) labeled with ruthenium complex 1.0 mg/L; phosphate buffer 50 mmol/L, pH 7.5; preservative.

Precautions and warnings

For in vitro diagnostic use.

Exercise the normal precautions required for handling all laboratory reagents.

Disposal of all waste material should be in accordance with local guidelines. Safety data sheet available for professional user on request.

Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).

Reagent handling

The reagents in the kit have been assembled into a ready-for-use unit that cannot be separated.

All information required for correct operation is available via the **cobas** link.

Storage and stability

Store at 2-8 $^{\circ}\text{C}.$

Do not freeze.

Store the **cobas e** pack **upright** in order to ensure complete availability of the microparticles during automatic mixing prior to use.



Stability:	
unopened at 2-8 °C	up to the stated expiration date
on the cobas e 801 analyzer	16 weeks

Specimen collection and preparation

Only the specimens listed below were tested and found acceptable. Serum collected using standard sampling tubes or tubes containing separating gel.

Li-heparin plasma. Do not use EDTA plasma.

Criterion: Recovery within \pm 30 % of serum value \geq 3.57 pmol/L (\geq 0.5 ng/mL); recovery within \pm 1.43 pmol/L (\pm 0.2 ng/mL) for serum value < 3.57 pmol/L (< 0.5 ng/mL) and slope of 0.9-1.1 + intercept within \pm 0.714 pmol/L (\pm 0.1 ng/mL) + coefficient of correlation \geq 0.95.

Stable for 3 hours at 20-25 °C, 5 days at 2-8 °C, 6 months at -20 °C (\pm 5 °C). Freeze only once.

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.

Centrifuge samples containing precipitates before performing the assay. Do not use heat-inactivated samples.

Do not use samples and controls stabilized with azide.

Ensure the samples and calibrators are at 20-25 °C prior to measurement.

Due to possible evaporation effects, samples and calibrators on the analyzers should be analyzed/measured within 2 hours.

Materials provided

See "Reagents – working solutions" section for reagents.

Materials required (but not provided)

- REF 07957203190, CalSet AMH Plus, for 4 x 1.0 mL
- REF 07957211190, PreciControl AMH Plus, for 4 x 2.0 mL
- REF 07299001190, Diluent Universal, 45.2 mL sample diluent
- General laboratory equipment
- cobas e 801 analyzer

Accessories for the cobas e 801 analyzer:

- REF 06908799190, ProCell II M, 2 x 2 L system solution
- REF 04880293190, CleanCell M, 2 x 2 L measuring cell cleaning solution
- REF 07485409001, Reservoir Cups, 8 cups to supply ProCell II M and CleanCell M
- REF 06908853190, PreClean II M, 2 x 2 L wash solution
- REF 05694302001, Assay Tip/Assay Cup tray, 6 magazines
 x 6 magazine stacks x 105 assay tips and 105 assay cups, 3 wasteliners
- REFJ 07485425001, Liquid Flow Cleaning Cup, 2 adaptor cups to supply ISE Cleaning Solution/Elecsys SysClean for Liquid Flow Cleaning Detection Unit
- REF 07485433001, PreWash Liquid Flow Cleaning Cup, 1 adaptor cup to supply ISE Cleaning Solution/Elecsys SysClean for Liquid Flow Cleaning PreWash Unit
- REF 11298500316, ISE Cleaning Solution/Elecsys SysClean, 5 x 100 mL system cleaning solution

Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

Resuspension of the microparticles takes place automatically prior to use.

Place the cooled (stored at 2-8 °C) **cobas e** pack on the reagent manager. Avoid foam formation. The system automatically regulates the temperature of the reagents and the opening/closing of the **cobas e** pack.

Calibration

Traceability: This method has been standardized against the Beckman Coulter AMH Gen II ELISA (unmodified version without predilution) assay.

The predefined master curve is adapted to the analyzer using the relevant CalSet.

Calibration frequency: Calibration must be performed once per reagent lot using fresh reagent (i.e. not more than 24 hours since the **cobas e** pack was registered on the analyzer). Renewed calibration is recommended as follows:

- after 12 weeks when using the same reagent lot
- after 28 days when using the same cobas e pack on the analyzer
- as required: e.g. quality control findings outside the defined limits

Quality control

For quality control, use PreciControl AMH Plus.

Additional suitable quality control material may also be used in combination with PreciControl AMH Plus.

Controls for the various concentration ranges should be run individually at least once every 24 hours when the test is in use, once per **cobas e** pack, and following each calibration.

Special care needs to be taken to ensure that the accuracy and precision of the testing stays within acceptable limits. Besides meeting the PreciControl AMH Plus target ranges provided, the user needs to ensure that the systematic bias with respect to the assigned target value is within \pm 12 %, the intermediate precision CV is \leq 8 % and the maximal total error is within \pm 25 % (TE = |biasl + 1.65*CV). It is recommended to use quality control rule software.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

If necessary, repeat the measurement of the samples concerned. Follow the applicable government regulations and local guidelines for quality control.

Calculation

The analyzer automatically calculates the analyte concentration of each sample (either in pmol/L or in ng/mL).

Conversion factors: $pmol/L \times 0.14 = ng/mL$ $ng/mL \times 7.14 = pmol/L$

Limitations - interference

The effect of the following endogenous substances and pharmaceutical compounds on assay performance was tested. Interferences were tested up to the listed concentrations and no impact on results was observed.

Endogenous substances

Compound	Concentration tested
Bilirubin	≤ 1129 µmol/L or ≤ 66 mg/dL
Hemoglobin	≤ 0.62 mmol/L or ≤ 1000 mg/dL
Intralipid	≤ 1000 mg/dL
Biotin	≤ 143 nmol/L or ≤ 30 ng/mL
IgG	≤ 2.5 g/dL
IgA	≤ 1.8 g/dL
IgM	≤ 0.5 g/dL

Criterion: Recovery within ± 10 % of initial value.

Samples should not be taken from patients receiving therapy with high biotin doses (i.e. > 5~mg/day) until at least 8 hours following the last biotin administration.

No interference was observed from rheumatoid factors up to a concentration of 1000 IU/mL.

There is no high-dose hook effect at AMH concentrations up to 9996 pmol/L (1400 ng/mL).



Pharmaceutical substances

In vitro tests were performed on 20 commonly used pharmaceuticals. No interference with the assay was found up to the concentrations indicated within the below table.

Active agent	Concentration tested mg/L
Acetylcysteine	1660
Ampicillin-Na	1000
Ascorbic acid	300
Cyclosporine	5
Cefoxitin	2500
Heparin	5000 U
Levodopa	20
Methyldopa	20
Metronidazole	200
Phenylbutazone	400
Doxycycline	50
Acetylsalicylic acid	1000
Rifampicin	60
Acetaminophen	200
Ibuprofen	500
Theophylline	100
Triptorelin acetate	0.1
Metformin	2000
Folic Acid	0.4
Levothyroxine	0.2

In rare cases, interference due to extremely high titers of antibodies to analyte-specific antibodies, streptavidin or ruthenium can occur. These effects are minimized by suitable test design.

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

Limits and ranges

Measuring range

0.07-164 pmol/L (0.01-23 ng/mL) (defined by the Limit of Detection and the maximum of the master curve). Values below the Limit of Detection are reported as <0.07 pmol/L (<0.01 ng/mL). Values above the measuring range are reported as >164 pmol/L (>23 ng/mL) or up to 328 pmol/L (46 ng/mL) for 2-fold diluted samples.

Lower limits of measurement

Limit of Blank, Limit of Detection and Limit of Quantitation

Limit of Blank = 0.049 pmol/L (0.007 ng/mL)

Limit of Detection = 0.07 pmol/L (0.01 ng/mL)

Limit of Quantitation = 0.214 pmol/L (0.03 ng/mL)

The Limit of Blank, Limit of Detection and Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A2 requirements.

The Limit of Blank is the 95^{th} percentile value from $n \ge 60$ measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95° %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples. The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

The Limit of Quantitation is the lowest analyte concentration that can be reproducibly measured with an intermediate precision CV of \leq 20 %.

Dilution

Samples with AMH concentrations above the measuring range can be diluted with Diluent Universal. The recommended dilution is 1:2 (either automatically by the analyzer or manually). The concentration of the diluted sample must be > 71.4 pmol/L (> 10 ng/mL).

After manual dilution, multiply the result by the dilution factor.

After dilution by the analyzers, the software automatically takes the dilution into account when calculating the sample concentration.

Expected values

A study in a Caucasian population with the Elecsys AMH Plus assay on samples from apparently healthy adults (148 males, 887 females not taking contraceptives) and 149 women with PCOS yielded the following results (Roche study No. RD001727):

	N	2.5 th perc.	5 th perc.	Median	95th perc.	97.5 th
		ng/mL	ng/mL	ng/mL	ng/mL	perc.
		(95 % CI ^{b)})	(95 % CI)	(95 % CI)	(95 % CI)	ng/mL
						(95 % CI)
Healthy	men					1
	148	0.77	1.43	4.79	11.6	14.5
		(0.17-1.58)	(0.256-1.97)	(4.35-5.35)	(10.3-17.0)	(10.9-17.6)
Healthy	wome	en (years)			•	ı
• 20-24	150	1.22	1.52	4.00	9.95	11.7
		(0.478-1.67)	(0.758-1.81)	(3.60-4.44)	(7.87-13.6)	(9.11-15.7)
• 25-29	150	0.890	1.20	3.31	9.05	9.85
		(0.493-1.21)	(0.797-1.75)	(3.00-3.89)	(7.59-10.3)	(8.91-11.3)
• 30-34	138	0.576	0.711	2.81	7.59	8.13
		(0.256-0.958)	(0.256-1.12)	(2.35-3.47)	(6.84-9.52)	(7.27-9.72)
• 35-39	138	0.147	0.405	2.00	6.96	7.49
		(0.053-0.474)	(0.053-0.496)	(1.73-2.36)	(5.31-9.37)	(6.49-10.9)
• 40-44	142	0.027	0.059	0.882	4.44	5.47
		(0.010-0.063)	(0.017-0.119)	(0.726-1.13)	(2.94-5.56)	(3.92-6.76)
• 45-50	169	0.010	0.010	0.194	1.79	2.71
		(0.010-0.010)	(0.010-0.010)	(0.144-0.269)	(1.43-2.99)	(1.79-4.16)
PCOS v	vomer	1*			ı	ı
	149	1.86	2.41	6.81	17.1	18.9
		(1.54-2.50)	(1.67-3.01)	(6.30-7.42)	(13.3-20.3)	(16.0-21.1)

b) CI = confidence interval

	N	2.5 th perc. pmol/L (95 % CI)	5 th perc. pmol/L (95 % CI)	Median pmol/L (95 % CI)	95 th perc. pmol/L (95 % CI)	97.5 th perc. pmol/L (95 % CI)	
Healthy	men						
	148	5.5	10.2	34.2	82.8	103	
		(1.2-11.3)	(1.8-14.1)	(31.1-38.2)	(73.5-121)	(78.1-125)	
Healthy	Healthy women (years)						
• 20-24	150	8.71	10.9	28.6	71.0	83.6	
		(3.41-11.9)	(5.41-12.9)	(25.7-31.7)	(56.2-97.1)	(65.0-112)	



	N	2.5th perc. pmol/L (95 % CI)	5 th perc. pmol/L (95 % CI)	Median pmol/L (95 % CI)	95 th perc. pmol/L (95 % CI)	97.5 th perc. pmol/L (95 % CI)
• 25-29	150	6.35	8.57 (5.69-12.5)	23.6 (21.4-27.8)	64.6 (54.2-73.5)	70.3 (63.6-81.0)
• 30-34	138	4.11 (1.83-6.84)	5.08 (1.83-8.00)	20.0 (16.8-24.8)	54.2 (48.8-68.0)	58.0 (51.9-69.4)
• 35-39	138	1.05 (0.378-3.38)	2.89 (0.378-3.54)	14.2 (12.4-16.9)	49.7 (37.9-66.9)	53.5 (46.3-77.9)
• 40-44	142	0.193	0.421 (0.121-0.850)	6.29 (5.18-8.07)	31.7 (21.0-39.7)	39.1 (28.0-48.3)
• 45-50	169	0.071	0.071	1.39 (1.03-1.92)	12.8 (10.2-21.3)	19.3
PCOS w	vomer	1*				
	149	13.3 (11.0-17.8)	17.2 (11.9-21.5)	48.6 (45.0-53.0)	122 (95.0-145)	135 (114-151)

^{*} According to the revised diagnostic criteria of PCOS defined by the Rotterdam ESHRE/ASRM-sponsored (ESHRE = European Society of Human Reproduction and Embryology; ASRM = American Society of Reproductive Medicine) PCOS consensus workshop group.²⁵

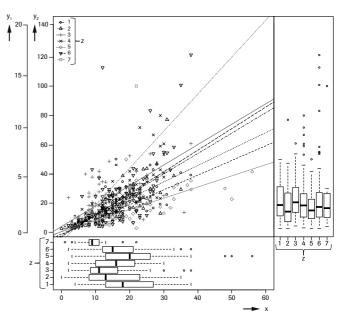
Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

Use of AMH for the assessment of ovarian reserve

The use of AMH for the assessment of ovarian reserve was investigated in a prospective study with n=451 women between 18-44 years old, where AMH values were correlated to the AFC of the women (Roche study No. RD001542). AFC was determined by transvaginal sonography measuring follicles of 2-10 mm diameter in size. Both AFC and AMH were determined on days 2-4 of the same menstrual cycle. Between 17 to 115 women were recruited per site at 6 different European sites and 1 Australian site.

No significant differences in mean AMH values were observed between the sites (pval = 0.301). The mean age values between the sites were significantly different, and also AMH and age showed a significant negative correlation (Spearman correlation coefficient -0.47). The age adjusted site effect of AMH showed no significance (pval = 0.193). The determined AFC values showed significant differences between the sites, with and without age adjustment. The overall correlation of AMH with AFC was 0.68 (Spearman's rank coefficient).

The figure below shows the scatterplot of AMH versus AFC, as well as the site-specific AMH and AFC distributions.



x: AFC (N)

y₁: AMH (ng/mL)

y₂: AMH (pmol/L)

z: Site

Agreement table on absolute AFC numbers of 7 and 15

Three AFC groups were defined 26,27 based on two cutoffs for AFC: 7 and 15 (0-7, 8-15, > 15). According to the prevalences within these groups (15 %, 37 %, 48 %), quantiles on AMH were computed (c1 = 4.86 pmol/L or 0.681 ng/mL, c2 = 16.2 pmol/L or 2.27 ng/mL) to define three groups. Agreement is presented in absolute numbers and percentages per AMH group.

Given the large variability of AFC results depending on site- and sonographer-specific variations, each site should review the agreement table for transferability to their own specific conditions.

	AFC 0-7	AFC 8-15	AFC > 15	N
AMH ≤ 4.86 pmol/L (0.681 ng/mL)	43 (63.2 %)	22 (32.4 %)	3 (4.4 %)	68
4.86 pmol/L (0.681 ng/mL) < AMH ≤ 16.2 pmol/L (2.27 ng/mL)	20 (12.0 %)	95 (56.9 %)	52 (31.1 %)	167
AMH > 16.2 pmol/L (2.27 ng/mL)	3 (1.4 %)	52 (24.1 %)	161 (74.5 %)	216
N	66	169	216	451

For a patient with AMH \leq 0.681 ng/mL the probability to have a low AFC (0-7) is 63 %, the probability to be in the middle AFC (8-15) group is about 32 % and only 4.4 % for having an AFC > 15.

The probability for patients with high AMH values (> 2.27 ng/mL) to have an AFC > 15 is 75 %, the probability for being in the middle AFC (8-15) group is 24 % and only 1.4 % to have an AFC < 8.

Use of AMH for the prediction of hyper-response to controlled ovarian stimulation

The following results were obtained in an external study "Clinical evaluation of the Elecsys AMH assay for the prediction of response to controlled ovarian stimulation" (Roche study No. CIM RD 001695).

AMH was determined in 149 women undergoing an antagonist treatment protocol in the course of their first cycle of controlled ovarian stimulation for in-vitro fertilization (IVF). Women included in the study were aged < 44 years, had a regular menstruation cycle and no major abnormalities at transvaginal sonography. Women with PCOS, endocrine or metabolic abnormalities and women undergoing IVF with oocyte donation were not



included. All women received a standard FSH stimulation dose of 150 IU/day. Blood was drawn before start of FSH stimulation for post hoc analysis of AMH after completion of the treatment cycle. Hyper-response was observed in 16 women. Hyper-response was defined as > 15 oocytes retrieved or cancellation of stimulation cycle where more than 20 follicles > 12 mm were observed and estradiol values > 11700 pmol/L, or when more than 30 follicles > 12 mm were observed. The clinical performance of Elecsys AMH to predict hyper-response to controlled ovarian stimulation was evaluated by ROC (receiver operating characteristic) analysis and by applying a cutoff of 15 pmol/L (2.1 ng/mL) which has been previously published. ^{28,29} Prediction of hyper-response was significant with an AUC (area under the curve) of 82.1 % (CI 72.5-91.7 %). Sensitivity, specificity, positive predictive value (PPV), and negative predictive value (NPV) for the AMH cutoff of 15.0 pmol/L (2.1 ng/mL) are shown in the table below.

Hyper-response							
AMH cutoff 15.0 pmol/L (2.10 ng/mL)							
	Estimate	95 % CI					
Sensitivity	81.3 %	54.4-96.0 %					
Specificity	64.7 %	55.9-72.8 %					
PPV	21.7 %	12.1-34.2 %					
NPV	96.6 %	90.5-99.3 %					

Use of AMH for individual daily dose determination of follitropin delta of Ferring

Follitropin delta is produced in a human cell line (PER.C6®) by recombinant DNA technology. ^{29,30,31} Ferring is marketing authorisation holder of follitropin delta.

The AMH concentration (in pmol/L) determined by the Elecsys AMH Plus assay in combination with body weight was validated for the individual daily dose determination of follitropin delta in controlled ovarian stimulation for the development of multiple follicles in women undergoing an assisted reproductive technology program such as an IVF or intracytoplasmic sperm injection (ICSI) cycle. The Elecsys AMH Plus assay was assessed exclusively for the individual dose determination of follitropin delta (Ferring). The AMH-based individualised dosing regimen of follitropin delta was validated in the prospective phase III clinical study ESTHER-1, a randomised, controlled, assessor-blind, parallel groups, multicenter, multinational trial comparing the efficacy and safety of follitropin delta with follitropin alfa (randomized in 1:1 ratio). Women aged 18-40 years undergoing controlled ovarian stimulation for IVF or ICSI were enrolled and followed a GnRH antagonist protocol. 32 In this trial, 665 IVF/ICSI patients randomised to the follitropin delta group were treated with an individual dose of follitropin delta determined on the basis of their body weight and their AMH (in pmol/L) concentration measured with the Elecsys AMH Plus assay. The individual daily dose of follitropin delta was maintained throughout stimulation with no dose adjustments.

Important information: Clinicians who want to administer follitropin delta must read and understand the current prescribing information of follitropin delta (Ferring) applicable in their country prior to administration of the drug; this document provides the details on the actual dosing regimen, the clinical efficacy and safety of the drug.³³

Specific performance data

Representative performance data on the analyzer is given below. Results obtained in individual laboratories may differ.

Precision

Precision was determined using Elecsys reagents, samples and controls in a protocol (EP05-A3) of the CLSI (Clinical and Laboratory Standards Institute): 2 runs per day in duplicate each for 21 days (n = 84). The following results were obtained:

cobas e 801 analyzer						
		Repeatability		Intermediate precision		
Sample	Mean pmol/L	SD pmol/L	CV %	SD pmol/L	CV %	
Human serum 1	0.307	0.007	2.4	0.011	3.6	
Human serum 2	6.50	0.098	1.5	0.151	2.3	

cobas e 801 analyzer						
	Repeatability		Intermediate precision			
Sample	Mean pmol/L	SD pmol/L	CV %	SD pmol/L	CV %	
Human serum 3	41.1	0.477	1.2	0.850	2.1	
Human serum 4	112	1.66	1.5	3.33	3.0	
Human serum 5	142	2.03	1.4	3.41	2.4	
PreciControl AMH 1	6.44	0.070	1.1	0.109	1.7	
PreciControl AMH 2	33.6	0.391	1.2	0.546	1.6	

cobas e 801 analyzer						
		Repeatability		Intermediate precision		
Sample	Mean ng/mL	SD ng/mL	CV %	SD ng/mL	CV %	
Human serum 1	0.0430	0.001	2.4	0.002	3.6	
Human serum 2	0.910	0.014	1.5	0.021	2.3	
Human serum 3	5.76	0.067	1.2	0.119	2.1	
Human serum 4	15.7	0.232	1.5	0.466	3.0	
Human serum 5	19.9	0.284	1.4	0.478	2.4	
PreciControl AMH 1	0.902	0.010	1.1	0.015	1.7	
PreciControl AMH 2	4.71	0.055	1.2	0.077	1.6	

Method comparison

A comparison of the Elecsys AMH Plus assay, REF 07957246190 (**cobas e** 801 analyzer; y) with the Elecsys AMH Plus assay, REF 07957190190 (**cobas e** 601 analyzer; x) gave the following correlations (ng/mL):

Number of samples measured: 126

Passing/Bablok³⁴ Linear regression y = 0.980x - 0.001 y = 0.951x + 0.100t = 0.979 r = 0.999

The sample concentrations were between 0.0117 and 22.1 ng/mL.

Analytical specificity

The monoclonal antibodies used are highly specific to human AMH. The following cross-reactivities were found:

Cross-reactant	Concentration tested	Cross-reactivity %
Inhibin A	100 ng/mL	n. d. ^{c)}
Activin A	100 ng/mL	n. d.
LH	500 mIU/mL	n. d.
FSH	500 mIU/mL	n. d.

c) n. d. = not detectable

References

- 1 Wilson CA, di Clemente N, Ehrenfels C, et al. Mullerian inhibiting substance requires its N-terminal domain for maintenance of biological activity, a novel finding within the transforming growth factor-beta superfamily. Mol Endocrinol 1993;7(2):247-257.
- 2 di Clemente N, Jamin SP, Lugovskoy A, et al. Processing of antimullerian hormone regulates receptor activation by a mechanism distinct from TGF-beta. Mol Endocrinol 2010;24(11):2193-206.
- Grinspon RP, Rey RA. Anti-müllerian hormone and sertoli cell function in paediatric male hypogonadism. Horm Res Paediatr 2010;73(2):81-92.



- 4 Broekmans FJ, Visser JA, Laven JS, et al. Anti-Müllerian hormone and ovarian dysfunction. Trends Endocrinol Metab 2008;19(9):340-347.
- Visser JA, de Jong FH, Laven JS, et al. Anti-Müllerian hormone: a new marker for ovarian function. Reproduction 2006;131(1):1-9.
- 6 Visser JA, Schipper I, Laven JS, et al. Anti-Müllerian hormone: an ovarian reserve marker in primary ovarian insufficiency. Nat Rev Endocrinol 2012;8(6):331-341.
- 7 Kelsey TW, Anderson RA, Wright P, et al. Data-driven assessment of the human ovarian reserve. Mol Hum Reprod 2012;18(2):79-87.
- 8 Nelson SM, Iliodromiti S, Fleming R, et al. Reference range for the antimüllerian hormone Generation II assay: a population study of 10,984 women, with comparison to the established Diagnostics Systems Laboratory nomogram. Fertil Steril 2014;101(2):523-529.
- 9 Tsepelidis S, Devreker F, Demeestere I, et al. Stable serum levels of anti-Mullerian hormone during the menstrual cycle: a prospective study in normo-ovulatory women. Hum Reprod 2007;22:1837–1840.
- 10 van Disseldorp J, Lambalk CB, Kwee J, et al. Comparison of inter- and intra-cycle variability of anti-Mullerian hormone and antral follicle counts. Hum Reprod 2010;25(1):221-227.
- 11 Overbeek A, Broekmans FJ, Hehenkamp WJ, et al. Intra-cycle fluctuations of anti-Müllerian hormone in normal women with a regular cycle: a re-analysis. Reprod Biomed Online 2012;24(6):664-669.
- 12 Kallio S, Puurunen J, Ruokonen A, et al. Antimüllerian hormone levels decrease in women using combined contraception independently of administration route. Fertil Steril 2013;99(5):1305-1310.
- 13 Nelson SM. Biomarkers of ovarian response: current and future applications. Fertil Steril 2013;99(4):963-969.
- 14 Anderson RA, Wallace WH. Antimüllerian hormone, the assessment of the ovarian reserve, and the reproductive outcome of the young patient with cancer. FertilSteril 2013;99(6):1469-1475.
- 15 Dewailly D, Andersen CY, Balen A, et al. The physiology and clinical utility of anti-Mullerian hormone in women. Hum Reprod Update 2014;20(3):370-385.
- 16 La Marca A, Sunkara SK. Individualization of controlled ovarian stimulation in IVF using ovarian reserve markers: from theory to practice. Hum Reprod Update 2014;20(1):124-140.
- 17 Josso N, Rey R, Picard JY. Testicular anti-Müllerian hormone: clinical applications in DSD. Semin Reprod Med 2012;30(5):364-373.
- 18 Rey RA, Grinspon RP, Gottlieb S, et al. Male hypogonadism: an extended classification based on a developmental, endocrine physiology-based approach. Andrology 2013;1(1):3-16.
- 19 La Marca A, Volpe A. The Anti-Mullerian hormone and ovarian cancer. Hum Reprod Update 2007;13(3):265-273.
- 20 Geerts I, Vergote I, Neven P, et al. The role of inhibins B and antimüllerian hormone for diagnosis and follow-up of granulosa cell tumors. Int J Gynecol Cancer 2009;19(5):847-855.
- 21 Dewailly D, Gronier H, Poncelet E, et al. Diagnosis of polycystic ovary syndrome (PCOS): revisiting the threshold values of follicle count on ultrasound and of the serum AMH level for the definition of polycystic ovaries. Hum Reprod 2011;26(11):3123-3129.
- 22 Iliodromiti S, Kelsey TW, Anderson RA, et al. Can anti-Mullerian hormone predict the diagnosis of polycystic ovary syndrome? A systematic review and meta-analysis of extracted data. J Clin Endocrinol Metab 2013;98(8):3332-3340.
- 23 Tehrani FR, Solaymani-Dodaran M, Tohidi M, et al. Modeling age at menopause using serum concentration of anti-mullerian hormone. J Clin Endocrinol Metab 2013;98(2):729-735.
- 24 Dólleman M, Depmann M, Eijkemans MJ, et al. Anti-Mullerian hormone is a more accurate predictor of individual time to menopause than mother's age at menopause. Hum Reprod 2014;29(3):584-591.
- 25 Rotterdam ESHRE/ASRM-Sponsored PCOS consensus workshop group. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome (PCOS). Hum Reprod 2004;19(1):41-47.

- 26 Ferraretti AP, La Marca A, Fauser BC, et al. ESHRE working group on Poor Ovarian Response Definition. ESHRE consensus on the definition of 'poor response' to ovarian stimulation for in vitro fertilization: the Bologna criteria. Hum Reprod 2011;26(7):1616-1624.
- 27 van Tilborg TC, Eijkemans MJ, Laven JS, et al. The OPTIMIST study: optimisation of cost effectiveness through individualised FSH stimulation dosages for IVF treatment. A randomised controlled trial. BMC Womens Health 2012;18;12:29.
- 28 Nelson SM, Yates RW, Fleming R. Serum anti-Müllerian hormone and FSH: prediction of live birth and extremes of response in stimulated cycles--implications for individualization of therapy. Hum Reprod 2007;22(9):2414-21.
- 29 Arce JC, Andersen AN, Fernández-Sánchez M, et al. Ovarian response to recombinant human follicle-stimulating hormone: a randomized, antimüllerian hormone-stratified, dose-response trial in women undergoing in vitro fertilization/intracytoplasmic sperm injection. Fertil Steril 2014;102(6):1633-40.e5.
- 30 Olsson H, Sandström R, Grundemar L. Different pharmacokinetic and pharmacodynamic properties of recombinant follicle-stimulating hormone (rFSH) derived from a human cell line compared with rFSH from a non-human cell line. J Clin Pharmacol. 2014;54(11):1299-307.
- 31 Bosch E, Nyboe Andersen A, Barri P, et al. Follicular and endocrine dose responses according to anti-Müllerian hormone levels in IVF patients treated with a novel human recombinant FSH (FE 999049). Clin Endocrinol (Oxf). 2015;83(6):902-12.
- 32 ClinicalTrials.gov Evidence-based Stimulation Trial with Human rFSH in Europe and Rest of World 1 (ESTHER-1). ClinicalTrials.gov Identifier: NCT01956110. Available at: http://clinicaltrials.gov/ct2/show/NCT01956110?term=NCT01956110.
- 33 Applicable prescribing information of follitropin delta; Ferring.
- 34 Bablok W, Passing H, Bender R, et al. A general regression procedure for method transformation. Application of linear regression procedures for method comparison studies in clinical chemistry, Part III. J Clin Chem Clin Biochem 1988 Nov;26(11):783-790.

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Symbols

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CONTENT Contents of kit

SYSTEM Analyzers/Instruments on which reagents can be used

REAGENT Reagent

CALIBRATOR Calibrator

Volume after reconstitution or mixing

GTIN Global Trade Item Number

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34 Bablok W, Passing H, Bender R, et al. A general regression procedure for method transformation. Application of linear regression procedures for method comparison studies in clinical chemistry, Part III. J Clin Chem Clin Biochem 1988 Nov;26(11):783-790.

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